

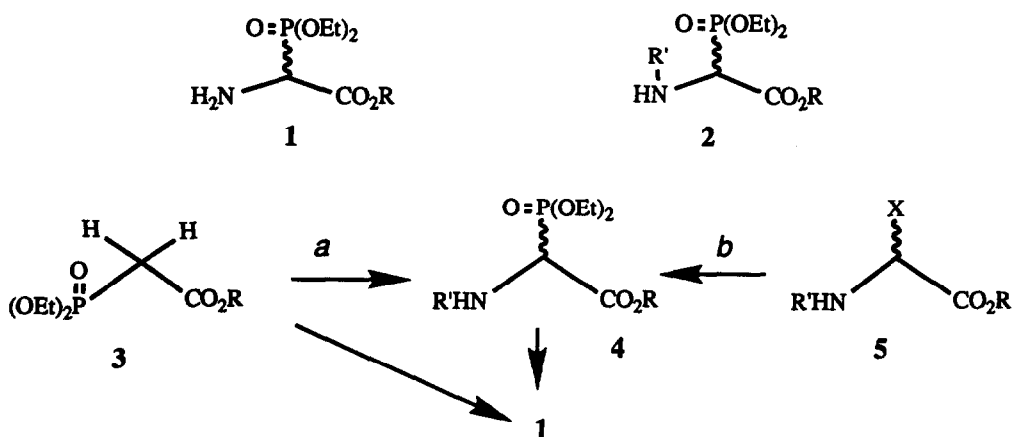
A Convenient Synthesis of 2-(Diethoxyphosphonyl)Glycine and its derivatives

R. Shankar and A. I. Scott*

Center for Biological NMR
Department of Chemistry, Texas A&M University
College Station, Tx. 77843

Abstract: A simple and very efficient method for the preparation of 2-(diethoxyphosphonyl)-glycine and its N-methyl analog *as the free acids* is described and their conversion to the corresponding t-butyl esters demonstrated.

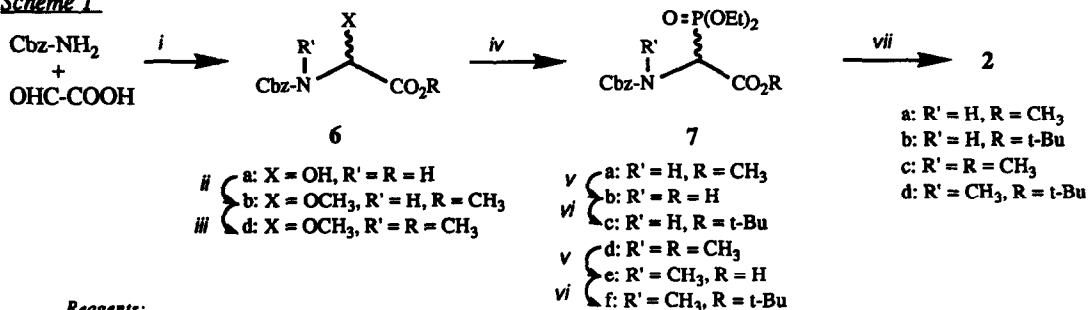
During the course of developing novel class of heterocycle-peptide conjugates and unique functionalized peptides as protease inhibitors¹, we required an efficient scheme for differential and selective protection of the carboxyl in 2-(diethoxyphosphonyl)glycine (1, R = Protecting group) and its N-alkyl derivatives (as 2). Originally introduced by Ratcliffe and Christensen² as the critical building block in the synthesis of (±) cephalothin, derivatives of 1 have been extensively employed in the construction of a large variety of β-lactam derivatives³⁻⁵, α-dehydro-α-amino acids and their oligopeptides⁶ and also are the subject of numerous patents.⁷



The preparation of 1 has been accomplished by direct amination of 2-(diethoxyphosphonyl) acetates⁸ (3, R = Me, Et or t-Bu; route a) or by Michaelis - Arbuzov reaction of N-protected α-halo glycine esters⁹ (5, X = Cl, Br; R = Me or Et, route b). However, in both

methods, the choice of carboxyl protection is restricted to the starting esters employed (route **a**) or limited to the stability of the protecting group to the preparative procedure (route **b**). The *t*-butyl group offers the best differential and orthogonal protection of the carboxyl functionality in peptide synthesis but, the only reported method¹⁰ for the preparation of such esters (**1**, R = *t*-Bu) is unsuitable on large scale due to the use of unstable and hazardous reagents coupled with low yields. The free acid **4** (R = H) should make the ideal starting material for the preparation of any desired ester of **1**. Although selective hydrolysis of the esters (as **4** prepared through either method) should furnish the free acid, no such preparations have been reported so far. Herein, we report a simple method for the preparation of **4** (R = H) and, for the first time, the *N*-methyl derivatives, **2** (R' = CH₃, R = H, CH₃ or *t*-Bu).

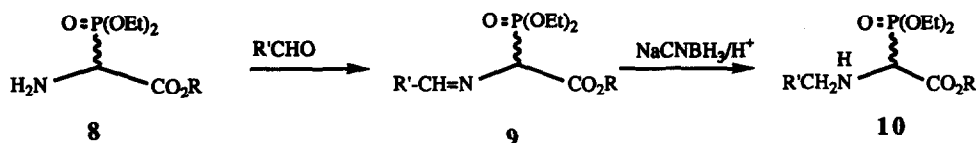
Scheme 1



Reagents:

i: ether, 16 h (95%); *ii*: MeOH/H₂SO₄, 48 h (98%); *iii*: MeI/NaH/DME/0→20°C, 16 h (98%); *iv*: PCl₅/CHCl₃/rt, 16 h; P(OEt)₃/70°C, 3 h (95%); *v*: 2M piperidine/rt, 6 h (100%); *vi*: *i*-butylene/H₂SO₄/CH₂Cl₂ (95%); *vii*: Pd/C/H₂ (42psi)/EtOH/HCl, 4 h (98%)

Scheme 2



N-Benzylloxycarbonyl- α -hydroxy glycine (**6a**) prepared from benzyl carbamate and glyoxylic acid was converted to methyl *N*-benzylloxycarbonyl-2-(diethoxyphosphonyl)glycinate (**7a**) by modification of the published procedure¹¹ in over 88% yield (Scheme 1). All attempts to selectively deprotect the carboxyl group in **7a** under a variety of basic and neutral conditions (LiOH/THF¹², K₂CO₃/MeOH¹³, EtSH/AlBr₃, LiI/Py¹⁴ etc.) resulted in inseparable reaction mixtures. Stronger acidic conditions also resulted in total decomposition of the ester. However, 2M piperidine in degassed water (5-7 eq.) effected quantitative hydrolysis of the methyl ester **7a**¹⁵ in 6 h at room temperature. The free acid **7b**¹⁶ was a stable solid with low solubility in most organic solvents at room temperature and could be converted to any desired carboxyl

derivative as demanded by the synthetic strategy employed. Thus, **7b** was converted to the *t*-butyl ester **7c**¹⁶ in over 95% isolated yield.

N-Alkylation of base-sensitive amines is often accomplished through the reduction of the corresponding Schiff's bases¹⁷ but, reduction of **9** (R' = Ph, R = CH₃) prepared from **8** (R = CH₃) and benzaldehyde (*Scheme 2*) gave an intractable mixture and was not investigated further. However, methyl N-benzyloxycarbonyl α -methoxy glycinate (**6b**) could be methylated directly with CH₃I/NaH¹⁸ to furnish **6d** in quantitative yield. Under these conditions, **6a** decomposed to benzyl carbamate and other unidentified products while **7a** underwent further methylation on the α -carbon. The methyl ester **7d**¹⁶ (prepared from **6d**) was saponified with 2M piperidine to **7e**¹⁶ and further transformed to the *t*-butyl ester **7f**¹⁶ (as before) in quantitative yield. Hydrogenolysis of the esters **7a**, **7c**, **7d** and **7f** gave the corresponding free amines **2a-d**¹⁶ in quantitative yield. This simple yet very efficient preparation of the free acids **7b** and **7e** offers ready access to any desired carboxyl derivative of this important building block in the synthesis of β -lactams, novel heterocycles, α -dehydro- α -amino acids and their oligopeptides.

A comment on the NMR spectra: ¹H and ¹³C spectra of the diethoxyphosphonates **7a-f** indicate that the two ethyl groups in the diethoxyphosphonyl moiety are non-equivalent, a fact further confirmed by molecular modelling studies. ³¹P-¹H and ³¹P-¹³C spin-coupling further contributes to the complexity of the NMR spectra of these compounds. Additionally, the N-methyl derivatives **6d** and **7d-f** exist as 3:1 mixtures of torsional isomers.

References and Notes

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15. Saponification of the corresponding ethyl ester was incomplete even after 48 h.
16. All the compounds were fully characterized (^1H and ^{13}C NMR, IR and MS). Data shown for selected products and only for the major isomer for compounds **7e-f**.
2b: ^1H NMR (500 MHz, CDCl_3) δ 1.27 and 1.29 (each t, $J = 7$ Hz, 3H), 1.43 (s, 9H), 1.79 (bs, 2H), 3.75 (d, $J = 19.8$ Hz, 1H), 4.15 (m, 4H); ^{13}C NMR (125 MHz, CDCl_3) δ 15.79, 27.21, 53.5 (d, $J = 138$ Hz), 62.29, 81.67 and 167.74; IR (film) 3400, 1735 and 1250 cm^{-1} ; M^+ (m/z): 267.
2c: ^1H NMR (500 MHz, CDCl_3) δ 1.26 and 1.28 (each t, $J = 7$ Hz, 6H), 2.28 (bs, 1H), 2.36 (s, 3H), 3.61 (d, $J = 23$ Hz, 1H), 3.75 (s, 3H) and 4.12 (m, 4H); ^{13}C NMR (125 MHz, CDCl_3) δ 15.86, 35.68, 52.1, 61.9 (d, $J = 147$ Hz), 63.04 (d, $J = 6$ Hz) and 169.27; M^+ (m/z): 239
2d: ^1H NMR (500 MHz, CDCl_3) δ 1.29 and 1.31 (each t, $J = 7$ Hz, 3H), 1.47 (s, 9H), 1.97 (bs, 1H), 2.4 (s, 3H), 3.47 (d, $J = 22$ Hz, 1H) and 4.15 (m, 4H); ^{13}C NMR (125 MHz, CDCl_3) δ 16.14, 27.67, 35.84, 62.66 (d, $J = 147.5$ Hz), 62.92 (d, $J = 6$ Hz), 82.23 and 168.0; IR (film) 3325, 1730 and 1255 cm^{-1} ; M^+ (m/z): 281
7b: ^1H NMR (500 MHz, CDCl_3) δ 1.19 and 1.20 (each t, $J = 6.8$ Hz, 3H), 4.09 and 4.15 (each m, 2H), 4.94 (dd, $J = 9.4, 22.9$ Hz, 1H), 5.12 (ABd, $J = 12.4$ Hz, 2H), 6.13 (d, $J = 9.4$ Hz, 1H) and 7.29 (m, 5H); ^{13}C NMR (125 MHz, CDCl_3) δ 15.9 and 15.93, 52.25 (d, $J = 150$ Hz), 64.14 and 64.38 (each d, $J = 6.3$ Hz), 67.16, 127.80-128.22, 135.8, 155.6 and 167.54; ^1H NMR (500 MHz, CD_3COCD_3) δ 1.25 (m, 6H), 4.14 (m, 4H), 4.85 (dd, $J = 9.4, 23.5$ Hz, 1H), 5.11 (ABd, $J = 12.4$ Hz, 2H), 6.9 (d, $J = 9.4$ Hz, 1H), 7.36 (m, 5H); ^{13}C NMR (125 MHz, CD_3COCD_3) δ 16.54, 53.5 (d, $J = 147$ Hz), 64.18, 67.29, 128.69-129.16, 137.26, 156.67 and 168.29; IR (KBr disk) 1740, 1695 and 1240 cm^{-1} ; M^+ (m/z): 345.
7c: m.p. 69° C (ether : hexane); ^1H NMR (500 MHz, CDCl_3) δ 1.17 and 1.21 (each t, $J = 6.8$ Hz, 3H), 1.37 (s, 9H), 4.02 (m, 4H), 4.68 (dd, $J = 9.16, 22.1$ Hz, 1H), 5.01 (ABd, $J = 12.2$ Hz, 2H), 5.74 (d, $J = 9.16$ Hz, 1H) and 7.22 (m, 5H); ^{13}C NMR (125 MHz, CDCl_3) δ 15.9, 27.47, 53.39 (d, $J = 146$ Hz), 63.12 (d, $J = 6.5$ Hz), 66.9, 83, 127.83-128.17, 135.79, 155.45 and 165.4; IR (KBr disk) 3240, 1750, 1720, 1235 cm^{-1} ; M^+ (m/z): 401.
7e: ^1H NMR (500 MHz, CDCl_3) δ 1.21 (m, 6H), 3.07 (s, 3H), 4.08 (m, 4H), 5.13 (ABd, $J = 12.2$ Hz, 2H), 5.54 (d, $J = 25.9$ Hz, 1H) and 7.3 (m, 5H); ^{13}C NMR (125 MHz, CDCl_3) δ 15.3, 20.6, 33.0, 57.0 (d, $J = 145$ Hz), 63.85 (d, $J = 6.5$ Hz), 67.8, 127-128.5, 135, 156.8 and 166.8; IR (film) 1740, 1705 and 1240 cm^{-1} ; M^+ (m/z): 359
7f: ^1H NMR (500 MHz, CDCl_3) δ 1.21 (m, 6H), 1.42 (s, 9H), 3.05 (s, 3H), 4.04 (m, 4H), 5.09 (ABd, $J = 12.2$ Hz, 2H), 5.36 (d, $J = 25.9$ Hz, 1H) and 7.25 (m, 5H); ^{13}C NMR (125 MHz, CDCl_3) δ 16.02, 27.64, 32.7, 57.6 (d, $J = 151$ Hz), 62.53 (d, $J = 6$ Hz), 67.54, 83.15, 127.5-128.21, 136.1, 156.6 and 165.09.
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